

Synthesis of purpurasol, a highly oxygenated coumarin from *Pterocaulon purpurascens*

Dominick Maes,^a Kris Van Syngel,^a Silvia Debenedetti^b and Norbert De Kimpe^{a,*}

^aDepartment of Organic Chemistry, Faculty of Bioscience Engineering, Ghent University, Coupure Links 653, B-9000 Ghent, Belgium

^bDepartment of Biological Sciences, Faculty of Sciences, National University of La Plata, Calle 115 and 47, 1900 La Plata, Argentina

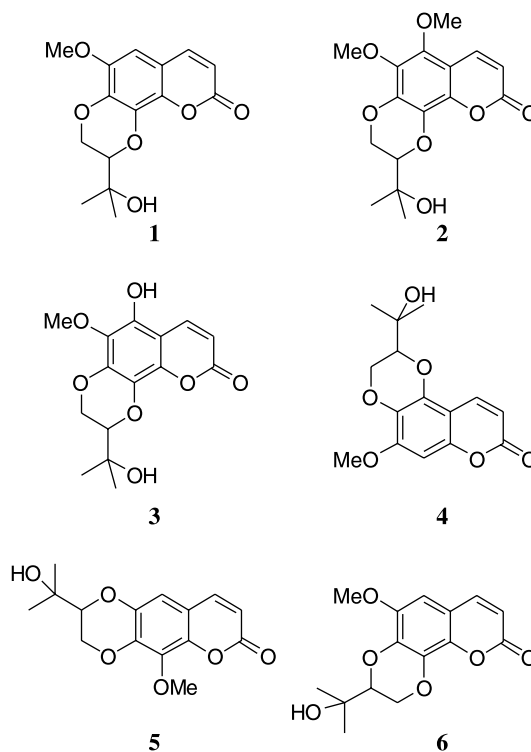
Received 23 January 2006; revised 13 February 2006; accepted 20 February 2006

Abstract—Purpurasol **1**, a 6,7,8-trioxygenated coumarin, isolated from *Pterocaulon purpurascens* (Asteraceae) and *Haplophyllum obtusifolium* (Rutaceae), was synthesized for the first time by a three-step synthesis starting from the natural coumarin fraxetin. This synthesis confirmed unambiguously the structure of purpurasol **1** and obtusifol.

© 2006 Elsevier Ltd. All rights reserved.

1. Introduction

The genus *Pterocaulon* is widely distributed in north eastern Argentina, southern Brazil and Paraguay. Plants of this genus are traditionally used in folk medicine for various purposes. The aerial parts of *Pterocaulon purpurascens* are used against snakebites and as an insecticide.^{1,2} A number of strongly related tri- and tetraoxygenated coumarins were isolated from *P. purpurascens*, more precisely purpureanol **2**,¹ purpurasol **1**² and purpurasolol **3**.³ Isopurpurasol **4**, a regioisomer of purpurasol, was isolated from another *Pterocaulon* species, namely *Pterocaulon virgatum*.⁴ All these coumarins show a characteristic benzodioxine moiety, which is very rare in natural coumarins.⁵ The present report deals with the synthesis of purpurasol **1** in order to secure unambiguously its structure and those of analogous coumarins **2** and **3**. The structure of purpurasol **1** was revealed based on spectroscopic data and by comparison with the earlier described purpureanol, of which the structure was unequivocally established based on X-ray spectroscopic analysis.¹ Later it was discovered that the spectroscopic, as well as the physical data obtained for purpurasol **1**, matched completely with those from an earlier described coumarin from *Haplophyllum obtusifolium*.^{6,7} The structure of this coumarin, which was given the trivial name obtusifol, was first proposed as **5**,⁶ but was later revised to **6**.^{7,8} Based on spectroscopic evidence it was shown that the coumarin from *H. obtusifolium* was identical to purpurasol **1**.⁹ In order to confirm this hypothesis, a synthesis of purpurasol was developed.



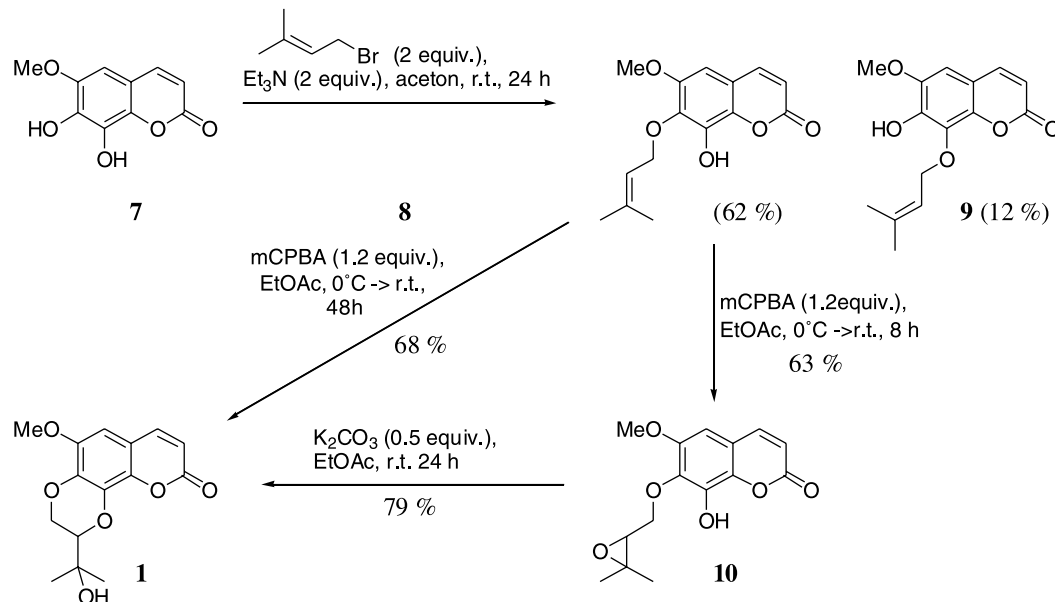
2. Results and discussion

Purpurasol **1** was synthesized starting from 7,8-dihydroxy-6-methoxy-2H-1-benzopyran-2-one, known as fraxetin **7**. Fraxetin **7** is a natural coumarin that occurs in many plant

Keywords: Coumarin; Purpurasol; *Pterocaulon purpurascens*.

* Corresponding author. Tel.: +32 9 264 59 51; fax: +32 9 264 62 43; e-mail: norbert.dekimpe@ugent.be

species, including *Aesculus turbinata* (Hippocastanaceae)¹⁰ and several *Fraxinus* spp. (Oleaceae).^{11,12} Fraxetin **7** was also isolated from *P. purpurascens*,³ which indicates that fraxetin **7** might be a natural precursor in the biosynthesis of purpurasol. The first synthetic step involved regioselective prenylation of fraxetin. Because of delocalization towards the electron withdrawing carbonyl group, the 7-OH group is



more acidic than the OH group at the 8-position. Although selective prenylation of fraxetin **7** was described before in 70% yield using sodium bicarbonate and 4-bromo-2-methyl-2-butene,¹³ in our hands only, and after numerous experiments under these and other reaction conditions, very low yields of the desired capensin **8** [8-hydroxy-6-methoxy-7-(3'-methyl-2'-butenyloxy)coumarin] were obtained. This problem of reproducibility is probably due to hydrolysis during aqueous workup. However, when the workup only consisted of the filtration of the reaction mixture, followed by rinsing the filter with dry acetone, the obtained yields were also very low. These low yields demanded for further evaluation of the reaction conditions. Much better results were obtained when the reaction was performed with triethylamine as a base. When fraxetin **7** was reacted with 2 equiv of prenylbromide and 2 equiv of triethylamine at room temperature for 24 h, the desired product, capensin **8** [8-hydroxy-6-methoxy-7-(3'-methyl-2'-butenyloxy)coumarin] was isolated in 62% yield. The regioisomeric 7-hydroxy-6-methoxy-8-(3'-methyl-2'-butenyloxy)coumarin **9** was isolated in 12% yield. In this reaction, purification was achieved by evaporating acetone from the reaction mixture and purifying the residual crude mixture by column chromatography. Capensin **8** is a naturally occurring coumarin and was isolated from several plant species, including *Phyllosma capensis*¹⁴ and *Bupleurum fruticosum*.¹⁵ To our knowledge 7-hydroxy-6-methoxy-8-(3'-methyl-2'-butenyloxy)coumarin **9** has never been reported from natural sources. The next step in the synthesis involved the epoxidation of the double bond of the prenyl group of capensin **8**. When capensin **8** was treated with 3-chloroperbenzoic acid in ethyl acetate, after 8 h 63%

of 7-(2,3-epoxy-3-methylbutoxy)-8-hydroxy-6-methoxycoumarin **10** was obtained. Cyclisation of epoxide **10** to purpurasol **1** was accomplished by treating it with potassium carbonate in ethyl acetate, affording purpurasol **1** in 79% yield. Purpurasol **1** could also be obtained in one step from capensin **8** by reaction with 3-chloroperbenzoic acid in ethyl acetate for 48 h, affording purpurasol **1** in 68% yield.

3. Experimental

3.1. General

¹H NMR spectra (300 MHz) and ¹³C NMR spectra (75 MHz) were recorded with a Joel Eclipse FT 300 NMR spectrometer. IR spectra were recorded on a Perkin Elmer Spectrum One spectrophotometer. Mass spectra were recorded on an Agilent 1100 Series VL mass spectrometer (ES 70 eV) or on a Varian MAT 112 mass spectrometer (EI 70 eV). Melting points were measured with a Büchi B-450 apparatus. Elemental analyses were measured with a Perkin-Elmer 2400 Elemental Analyzer. Flash chromatography was performed with ACROS silica gel (particle size 0.035–0.070 mm, pore diameter ca. 6 nm) using a glass column. 7,8-Dihydroxy-6-methoxycoumarin was obtained from Aldrich Chemical Company. All other reagents were obtained from Acros Organics and were used as such, except for 3-chloroperbenzoic acid. 3-Chloroperbenzoic acid ($\leq 77\%$, remainder 3-chlorobenzoic acid and water) was obtained from Acros Organics and was kept under reduced pressure (5 mmHg) at room temperature for 3 h in order to remove most of the water.

3.2. Synthetic procedures

3.2.1. 8-Hydroxy-6-methoxy-7-(3'-methyl-2'-butenyloxy)-coumarin (capensin) **8 and 7-hydroxy-6-methoxy-8-(3'-methyl-2'-butenyloxy)coumarin **9**.** 7,8-Dihydroxy-6-methoxycoumarin **7** (208 mg, 1 mmol) was dissolved in 10 ml acetone, and 202 mg (2 mmol) of triethylamine and 298 mg (2 mmol) of 4-bromo-2-methyl-2-butene were added.

After stirring at room temperature for 24 h, the acetone was evaporated in vacuo and the residue was chromatographed over silica gel using 40% hexane, 50% diethyl ether and 10% tetrahydrofuran as mobile phase. This procedure yielded 62% (172 mg) of 8-hydroxy-6-methoxy-7-(3'-methyl-2'-butenyloxy)coumarin (capensin) **8** ($R_f=0.23$) and 12% (21 mg) 7-hydroxy-6-methoxy-8-(3'-methyl-2'-butenyloxy)coumarin **9** ($R_f=0.14$), both appearing as a yellow powder.

8-Hydroxy-6-methoxy-7-(3'-methyl-2'-butenyloxy)-coumarin (capensin) 8

Mp (°C): 135 (lit. 135–136¹). IR (KBr, cm⁻¹): 3392 (broad, OH); 1692 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 1.68 (3H, s, 4'-CH₃); 1.75 (3H, s, 5'-CH₃); 3.90 (3H, s, OCH₃); 4.69 (1H, d, $J=7.4$ Hz, 1'-CH₂); 5.52 (1H, t, $J=7.4$ Hz, 2'-CH); 6.16 (1H, br s, OH); 6.34 (1H, d, $J=9.6$ Hz, 3-CH); 6.50 (1H, s, 5-CH); 7.62 (1H, d, $J=9.6$ Hz, 4-CH). ¹³C NMR (68 MHz, CDCl₃): δ 18.0 en 25.9 (2×CH₃); 56.2 (OCH₃); 70.0 (1'-CH₂); 100.0 (5-CH); 114.4 (C_q); 115.2 (3-CH); 119.5 (2'-CH); 137.8 (C_q); 138.0 (2×C_q); 140.5 (3'-C_q); 143.7 (4-CH); 149.8 (6-C_q); 160.4 (C=O). MS (70 eV, ES, m/z (%)): 275 (M-1). Anal. Calcd for C₁₅H₁₆O₅: C, 65.21%; H, 5.84%. Found: C, 65.01%; H, 5.68%.

7-Hydroxy-6-methoxy-8-(3'-methyl-2'-butenyloxy)-coumarin 9

Mp (°C): 126.5. IR (KBr, cm⁻¹): 3401 (broad, OH); 1702 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 1.70 (3H, s, 4'-CH₃); 1.75 (3H, s, 5'-CH₃); 3.93 (3H, s, OCH₃); 4.80 (1H, d, $J=7.4$ Hz, 1'-CH₂); 5.55 (1H, t, $J=7.4$ Hz, 2'-CH); 6.27 (1H, d, $J=9.6$ Hz, 3-CH); 6.66 (1H, s, 5-CH); 7.61 (1H, d, $J=9.6$ Hz, 4-CH). ¹³C NMR (75 MHz, CDCl₃): δ 18.1 en 25.9 (2×CH₃); 56.4 (OCH₃); 70.3 (1'-CH₂); 103.5 (5-CH); 111.1 (C_q); 113.3 (3-CH); 119.4 (2'-CH); 133.1 (C_q); 140.6 (C_q); 143.0 (C_q); 143.2 (C_q); 143.9 (4-CH); 144.6 (6-C_q); 160.7 (C=O). MS (70 eV, ES, m/z (%)): 275 (M-1). Anal. Calcd for C₁₅H₁₆O₅: C, 65.21%; H, 5.84%. Found: C, 65.39%; H, 5.98%.

3.2.2. 7-(2,3-Epoxy-3-methylbutoxy)-8-hydroxy-6-methoxycoumarin 10. 8-Hydroxy-6-methoxy-7-(3'-methyl-2'-butenyloxy)coumarin (capensin) **8** (69 mg, 0.25 mmol) was dissolved in 2.5 ml of ethyl acetate. The reaction mixture was cooled to 0 °C and 52 mg (0.3 mmol) of 3-chloroperoxybenzoic acid was added, after which the reaction was stirred at room temperature for 8 h. The solvent was evaporated in vacuo and the resulting mixture was dissolved in dichloromethane (25 ml) and washed with 25 ml of saturated aqueous sodium bicarbonate and 20 ml of water, respectively. The organic phase was dried (MgSO₄) and after filtration and evaporation of the solvent, 46 mg (63%) of crude 7-(2,3-epoxy-3-methylbutoxy)-8-hydroxy-6-methoxycoumarin **10** (purity: 95%) was obtained as a sticky residue, which was used without further purification.

IR (KBr, cm⁻¹): 3419 (broad, OH); 1715 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 1.32 en 1.38 (each 3H, each s, (CH₃)₂C=), 3.23 (1H, dd, $J=6.60$, 4.68 Hz, OCHCH₂O), 3.90 (3H, s, OCH₃), 4.16 (1H, dd, $J=6.60$, 11.56 Hz, OCHCH_aH_bO), 4.44 (1H, dd, $J=4.68$, 11.56 Hz, OCHCH_aH_bO), 6.35 (1H, d, $J=9.63$ Hz, 3-CH), 6.50 (1H, s, 5-CH),

7.62 (1H, d, 4-CH). ¹³C NMR (75 MHz, CDCl₃): δ 18.7 en 24.6 (2×CH₃); 56.1 (OCH₃); 59.3 (3'-C_q); 61.5 (2'-CH); 72.1 (1'-CH₂); 99.7 (5-CH); 114.6 (C_q); 115.2 (3-CH); 138.0 (2×C_q); 138.3 (C_q); 143.8 (4-CH); 149.8 (6-C_q); 160.6 (C=O). MS (70 eV, EI, m/z (%)): 292 (M⁺, 16); 208 (100); 193 (15); 139 (20); 85 (75); 83 (17); 71 (52); 69 (64); 55 (32); 49 (16); 43 (73).

3.2.3. Purpurasol 1.

Procedure 1

7-(2,3-Epoxy-3-methylbutoxy)-8-hydroxy-6-methoxycoumarin **10** (29 mg, 0.1 mmol) was dissolved in 1 ml of EtOAc. 0.05 mmol (7 mg) of potassium carbonate was added and the reaction was stirred at room temperature for 24 h. Water (10 ml) and ethyl acetate (5 ml) were added, and the aqueous phase was further extracted with 2×10 ml of ethyl acetate. The combined organic layers were dried (MgSO₄) and, after filtration and evaporation of the solvent, 23 mg (79%) of purpurasol was obtained as a white crystalline solid, which was recrystallized from ethanol.

Procedure 2

8-Hydroxy-6-methoxy-7-(3'-methyl-2'-butenyloxy)-coumarin (capensin) **8** (28 mg, 0.1 mmol) was dissolved in 1 ml of ethyl acetate. The reaction mixture was cooled to 0 °C and 20 mg (0.12 mmol) of 3-chloroperoxybenzoic acid was added. The reaction was stirred at room temperature for 48 h. The solvent was evaporated and the resulting mixture was dissolved in dichloromethane (10 ml) and washed with 10 ml of saturated aqueous sodium bicarbonate and 10 ml of water, respectively. The organic phase was dried (MgSO₄) and, after filtration and evaporation of the solvent, 20 mg (69%) of purpurasol was obtained as a white solid.

Mp (°C): 148 (lit. 148–149²). IR (KBr, cm⁻¹): 3400 (broad, OH); 1702 (C=O). ¹H NMR (300 MHz, CDCl₃): δ 1.37 en 1.46 (2×3H, s, CH₃); 2.75 (1H, br s, OH); 3.92 (3H, s, OCH₃); 3.99 (1H, dd, $J_{ax}=9.1$ Hz, $J_{bx}=1.9$ Hz, 2'-CH); 4.13 (1H, dd, $J_{ab}=11.3$ Hz, $J_{ax}=9.1$ Hz, 1'-CH_aH_b); 4.65 (1H, dd, $J_{ab}=11.3$ Hz, $J_{bx}=1.9$ Hz, 1'-CH_aH_b); 6.31 (1H, d, $J=9.6$ Hz, 3-CH); 6.51 (1H, s, 5-CH); 7.61 (1H, d, $J=9.6$ Hz, 4-CH). ¹³C NMR (75 MHz, CDCl₃): δ 25.1 en 26.0 (2×CH₃); 56.4 (OCH₃); 65.5 (1'-CH₂); 70.6 (3'-C_q); 79.0 (2'-C_q); 100.1 (5-CH); 111.6 (4a-C_q), 114.1 (3-CH); 132.4 (8-C_q); 136.7 (8a-C_q); 139.0 (7-C_q); 143.8 (4-CH); 145.7 (6-C_q); 160.9 (C=O). MS (70 eV, EI, m/z (%)): 293 (M+1, 35); 292 (M⁺, 99); 235 (26); 234 (100); 219 (57); 208 (46); 207 (23); 206 (23); 205 (78); 191 (23); 176 (25); 79 (35); 71 (21); 59 (92); 57 (52); 51 (26); 47 (23); 43 (75). Anal. Calcd for C₁₅H₁₆O₆: C, 61.64%; H, 5.52%. Found: C, 61.49%; H, 5.40%.

References and notes

- Debenedetti, S. L.; Nadinic, E. L.; Coussio, J. D.; De Kimpe, N.; Feneau-Dupont, J.; Declercq, J.-P. *Phytochemistry* **1991**, 2757–2758.
- Debenedetti, S. L.; Nadinic, E. L.; Gomez, M. A.; Coussio, J. D.; De Kimpe, N.; Boeykens, M. *Phytochemistry* **1992**, 3284–3285.

3. Debenedetti, S. L.; Nadinic, E. L.; Gomez, M. A.; Coussio, J. D.; De Kimpe, N.; Boeykens, M. *Phytochemistry* **1996**, 563–564.
4. Debenedetti, S. L.; Abbaspour Tehrani, K.; Van Puyvelde, L.; De Kimpe, N. *Phytochemistry* **1999**, 701–703.
5. Murray, R.D.H. *Fortschritte der Chemie organischer Naturstoffe—Progress in the Chemistry of Organic Natural Compounds*; Springer-Verlag: Vienna, 2002; Vol. 83.
6. Gashimov, N. F.; Kuznetsova, G. A. *Khim. Prir. Soedin.* **1974**, 303.
7. Abyshev, A. Z.; Gashimov, N. F. *Khim. Prir. Soedin.* **1979**, 401.
8. Abyshev, A. Z.; Zmeykov, V. P.; Sidorova, I. P. *Khim. Prir. Soedin.* **1982**, 301.
9. Boeykens, M.; De Kimpe, N.; Debenedetti, S. L.; Nadinic, E. L.; Gomez, M. A.; Coussio, J. D.; Abyshev, A. Z.; Gindin, V. A. *Phytochemistry* **1994**, 1559–1560.
10. Shimada, H. *Yakugaku Zasshi (J. Pharm. Soc. Jpn.)* **1937**, 148.
11. Shimada, H. *Yakugaku Zasshi (J. Pharm. Soc. Jpn.)* **1938**, 185.
12. Shimada, H. *Yakugaku Zasshi (J. Pharm. Soc. Jpn.)* **1952**, 63.
13. Ahluwalia, V. K.; Khanna, M.; Singh, R. P. *Gazz. Chim. Ital.* **1981**, 111, 503.
14. Campbell, W. E.; Gragg, G. M. L. *Phytochemistry* **1979**, 688–689.
15. Pistelli, L.; Bertoli, A.; Bilia, A. R.; Morelli, I. *Phytochemistry* **1996**, 1579–1582.